

- What is virus?
- Symptoms
- Transmission
- Detection and identification
- Management

Introduction

- What is virus?
 - one kind of plant pathogens
 - smaller than bacteria
 - not a cell... "particle"
 - can cause severe diseases
- composed mainly of nucleic acid and coat protein
- usually too small to be seen under light microscope
- Different shape and size; spherical, rod

Introduction

- More than 2,000 kinds of viruses are found and new viruses are described every month.
- ➤ One virus may infect one or dozens of different species of plants "host range"
- A plant may be infected with one or more of viruses.
- Propagate only in host cells (obligate parasite)
- > Some can propagate in insect
- > Enter plants by wounds or vectors
- not infect human or animals

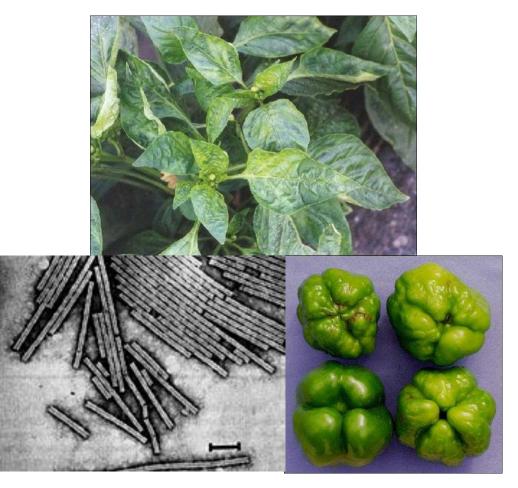
Can a plant virus make you sick?

By Vincent Racaniello, A virology Professor

- One hundred trillion bacteria colonize the human intestine, 10 times the numbers of cells in entire human body.
- The vast majority of DNA viruses in the human gut are bacteriophages, which infect the resident bacteria.
- The most plentiful RNA viruses in our feces are plant viruses, and the most abundant is pepper mild mottle virus (PMMV) up to 10⁹ virions per gram.
- http://www.virology.ws/2010/04/29/can-a-plant-virus-make-you-sick/

• High levels of PMMV are found in <u>Tabasco sauce</u>, which contains virions that are not only visible in the electron microscope, but which are infectious for plants.







Archives of Virology

-- August 2015, Volume 160, <u>Issue 8</u>, pp 2079–2082

Detection of pepper mild mottle virus in pepper sauce in China

Jiejun Peng, Bingbin Shi, Hongying Zheng, Yuwen Lu, Lin Lin, Tong Jiang, Jianping Chen ⊠, Fei Yan ⊠

Brief Report

First Online: 29 May 2015

DOI: 10.1007/s00705-015-

2454-7

Cite this article as:

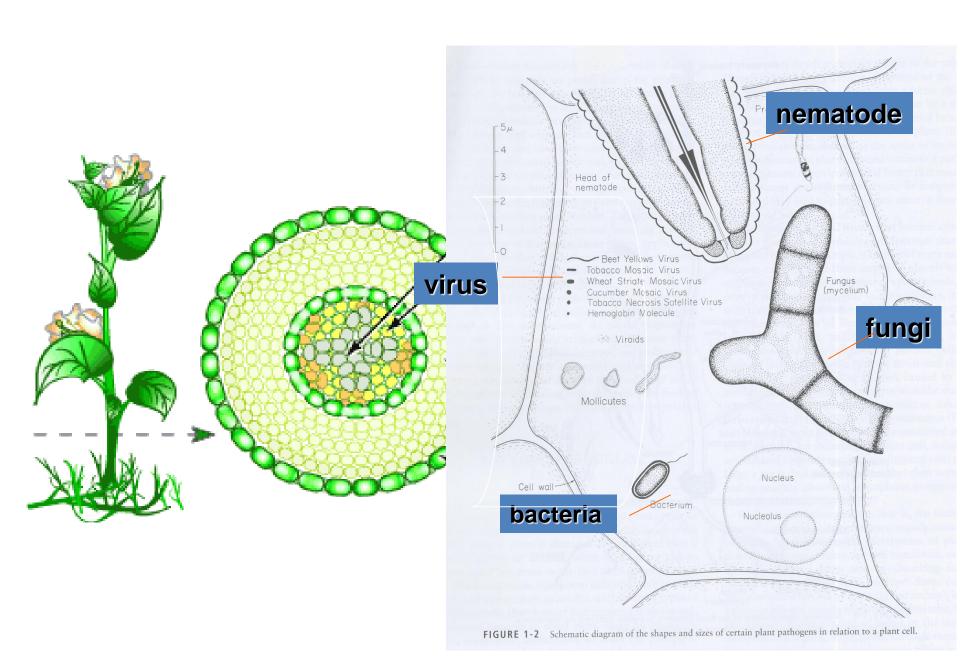
Peng, J., Shi, B., Zheng, H. et al. Arch Virol (2015) 160: 2079.

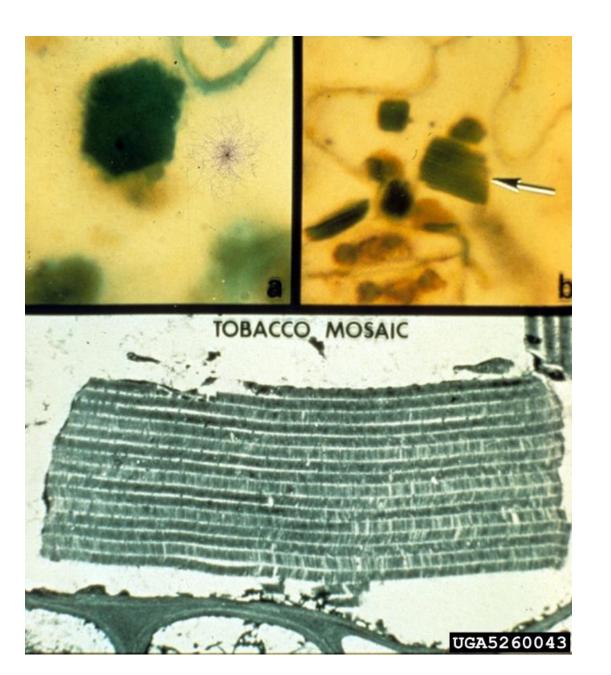
doi:10.1007/s00705-015-2454-7



Abstract

Pepper mild mottle virus (PMMoV) was detected by RT-PCR in all 42 pepper sauce samples from the 10 main manufacturing provinces in China at concentrations ranging from 3.8 to 8.8 (Log₁₀ copies/mL). Their coat protein nucleotide sequences had 97.4 to 100 % identity to each other and 92.4 to 100 % to other published isolates. The samples remained infectious to *N. benthamiana*, indicating that commercial trade in sauce could contribute to the natural spread of PMMoV.

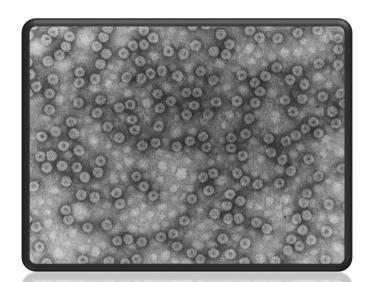


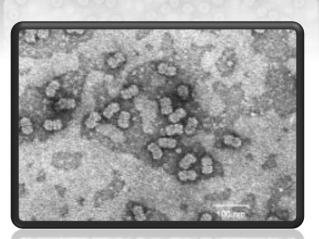


Inclusion bodies

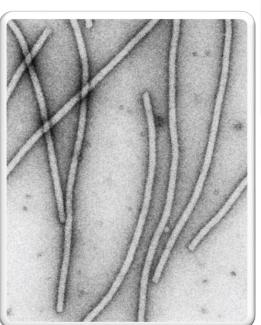
Tobacco mosaic virus is the first discovered plant virus

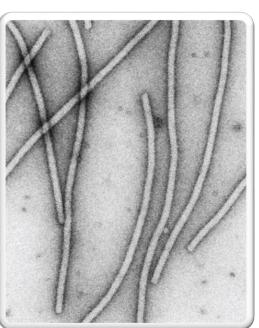
Morphology of plant viruses under TEM

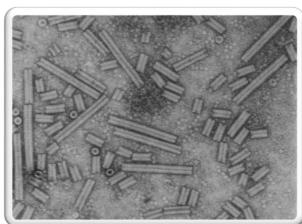


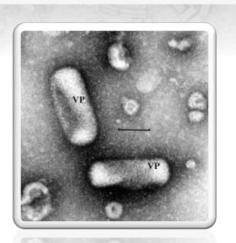


spherical



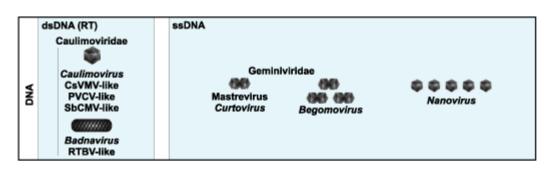


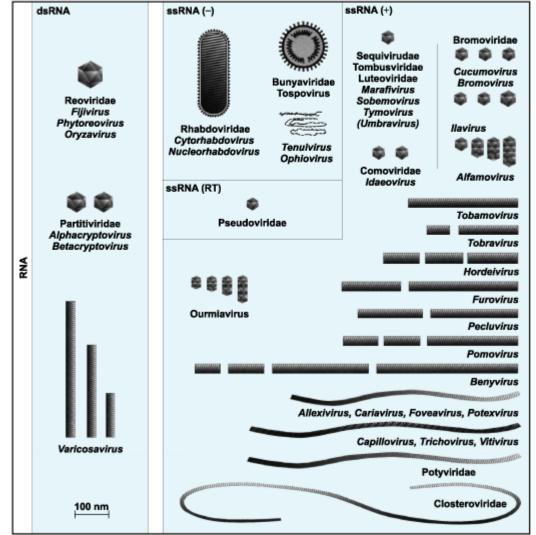




rod

Families and Genera of Viruses Infecting Plants





Nomenclature of plant virus

- Tobacco mosaic virus (TMV)
- Tomato yellow leaf curl virus (TYLCV)
- Bean yellow mosaic virus (BYMV)

(Plant + symptom + virus)

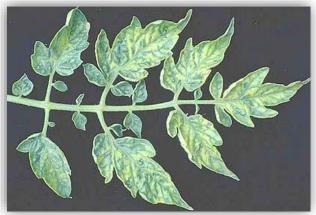
"Virus species"

Symptoms caused by plant viruses



Summer squash leaves show mosaic symptoms caused by *Watermelon Mosaic Potyvirus* (formerly WMV-2).





Tomato Mosaic Virus (ToMV)Strong yellow mosaic symptoms



orange - mottle



Orchid – mosaic flower



Gerbera - mosaic

Tulip - mosaic

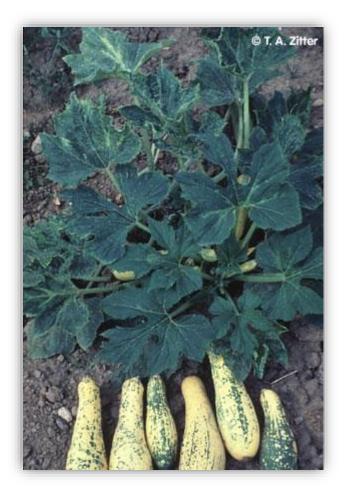




Melon fruit infected with **Squash Mosaic Comovirus** show strong mottled patterns and lack of netting



Cucumber Mosaic Virus (CMV)



Mosaic and rugosity of foliage and color breaking on fruit of straightneck yellow squash caused by Watermelon Mosaic Potyvirus (formerly WMV-2).

(rugose = wrinkle)





Common mosaic seed discoloration caused by **soybean mosaic virus**







Yellow vein mosaic virus

yellows-type disease on tomato plant caused by **phytoplasma**



Leaf yellowing on squash caused by Cucurbit aphidborne yellows virus





Corn - streak mosaic

Sugarcane - mosaic





Papaya ringspot virus (PRSV)

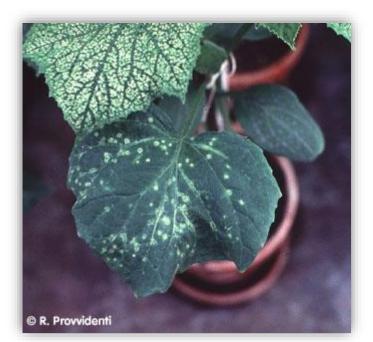


Severe blight by *maize chlorotic mottle virus* (MCMV)



Peronosclerospora maydis





Tobacco Ringspot Nepovirus
infections appear as yellowish
mosaic on newly infected leaves of
melon and cucumber





Papaya ringspot virus (PRSV)



Jalapeno peppers showing typical symptoms caused by PMMoV (left) and *Tomato* spotted wilt virus (TSWV) (right)



Turnip mosaic virus symptoms on the internal leaves of cabbage.



Tomato
Mosaic Virus
(ToMV)
Necrotic spots
on fruit



upward and inward rolling of the leaf margins





Tomato yellow leaf curl virus (TYLCV). Note multiple shoots, thickened shoots and deformed yellow

Several symptoms can be caused by one virus



Leaf mosaic



Water-soaked streak on stem



Ring spot on fruit



Leaf deformation

Several symptoms can be caused by one virus

Wide host range >40 families





Cucumber mosaic virus (CMV)





Wilt -Tobacco etch virus (TEV)



Reduced fruit size and deformation





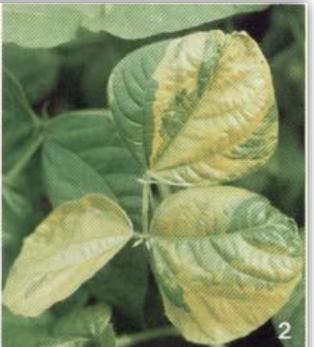
Bean yellow mosaic vrius

similar symptoms may caused by different viruses



Bean common mosaic virus





Virus-like diseases

genetics

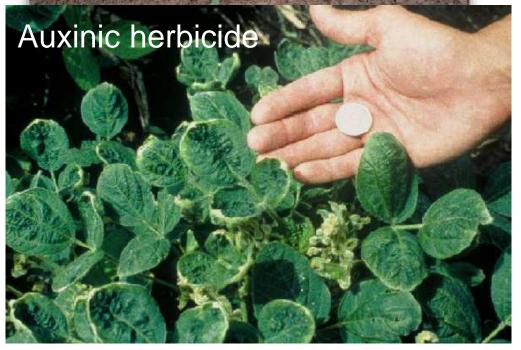


chemicals

2,4-D









Herbicides

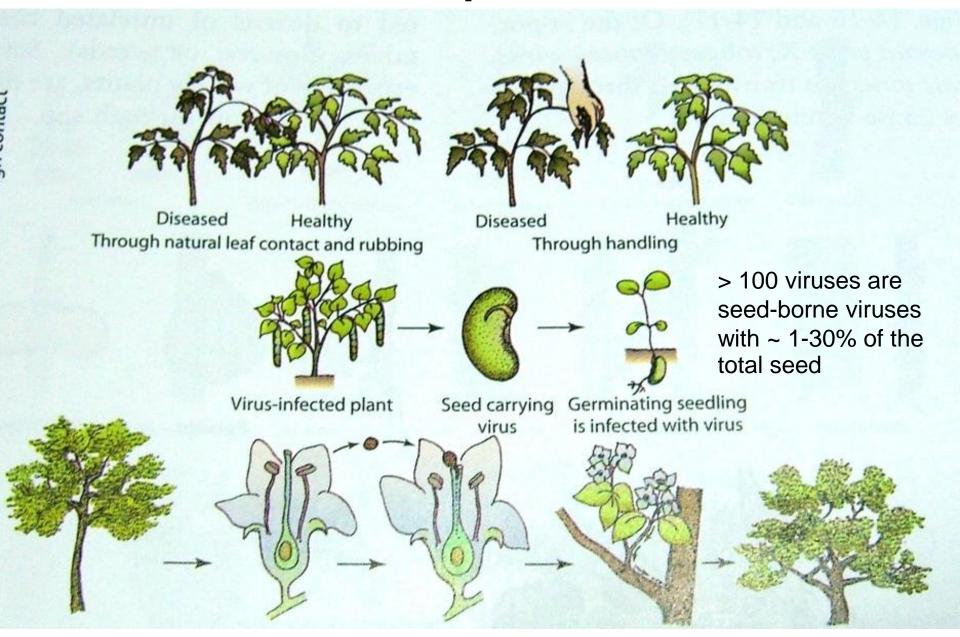


Hormone effect

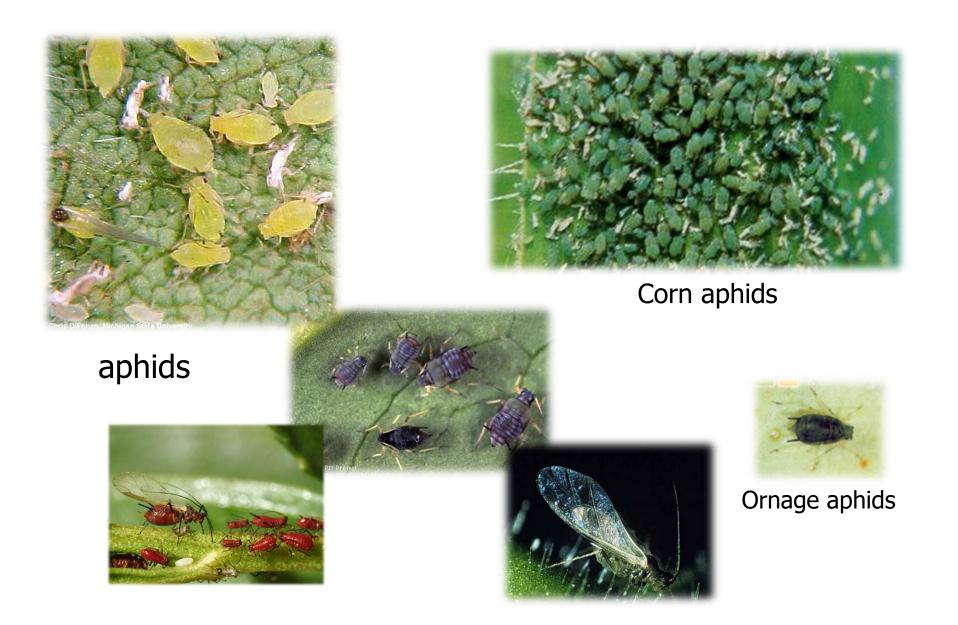


DISEASE TRANSMISSION

Mechanical, seed, pollen transmission



Insect transmission





White flies



thrip



hopper

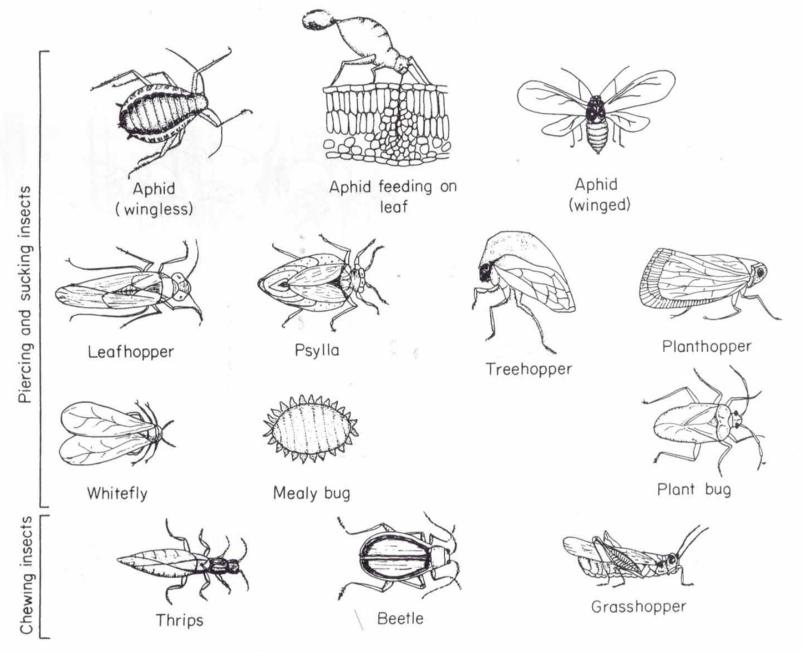


FIGURE 14-18 Insect vectors of plant viruses. Insects in second row from the top also transmit mollicutes and fastidious vascular bacteria.

Inoculativity = the ability of an aphid or other insect to deliver virus into a healthy plant.

Acquisition feed = the feeding process by which the insect acquires virus from an infected plant.

Inoculative feed = the feed during which virus is delivered into a healthy plant.

Latent period = the time after acquisition feed for which the aphid is unable to transmit the virus.

Retention time = how long the aphid retains the ability to infect plant.

Circulative transmission

(persistent)

Nonpropagative
Virus do not replicate in insect



Propagative
Replicate in aphid vector,

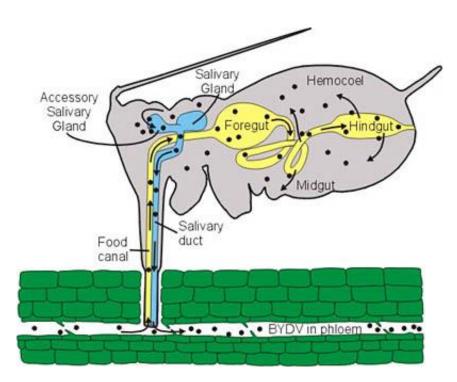
Noncirculative transmission

Nonpersistent

Stylet tip, food canal, foregut, short retention time

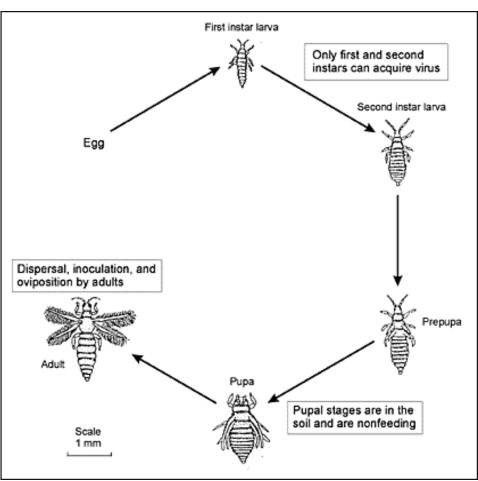
Semipersistent

Adsorbed to foregut, Virus confined to phloem, Longer acq. feed

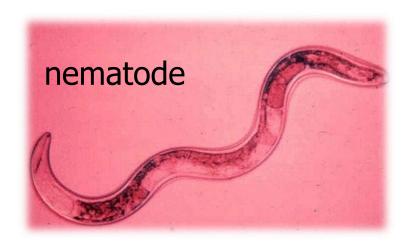


persistent transmission: (syn. circulative transmission)

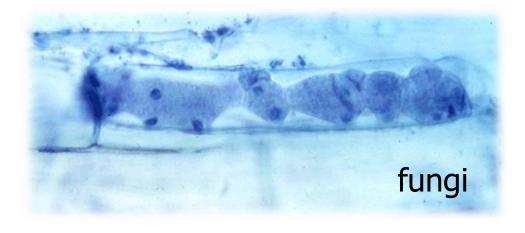
a type of virus transmission in which the virus is acquired and transmitted by the vector after relatively long feeding times and remains transmissible for a prolonged period while in association with its vector (aphid vector feeding on a plant host showing the internal route of the viruses that cause barley yellow dwarf)

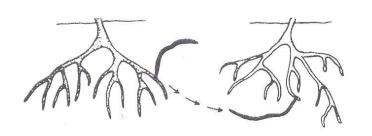


white fly life cycle





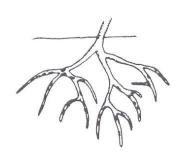




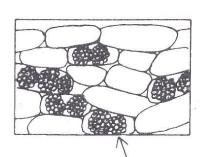
Virus transmission by nematodes

Eriophyid mite

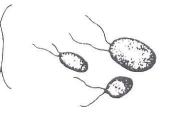
Mite vector of plant viruses



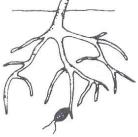
Plant infected with virus and fungus



Fungal zoòsporangia in root of virus-infected plant



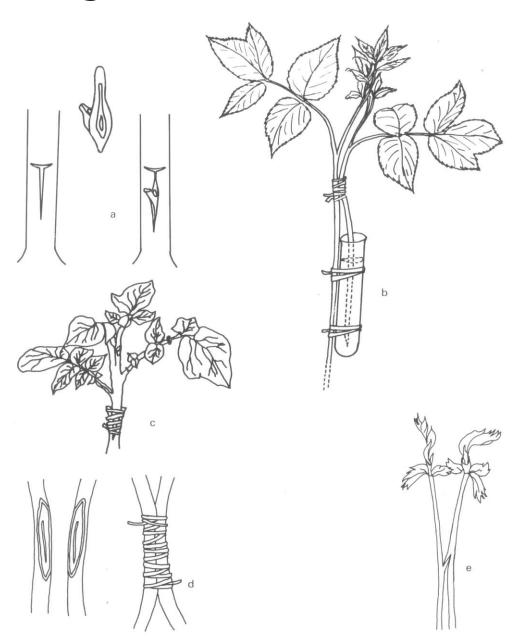
Virus-carrying zoospores leave plant



Zoospore infects new plant and transmits virus

FIGURE 14-20 Transmission of plant viruses by nematodes, mites, and fungi.

Vegetative transmission



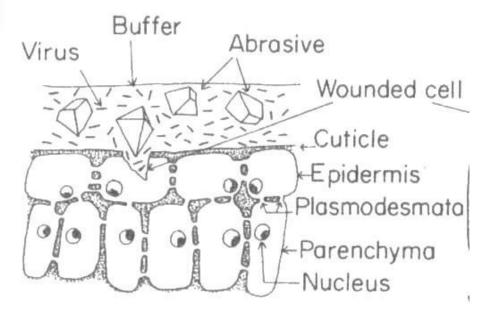


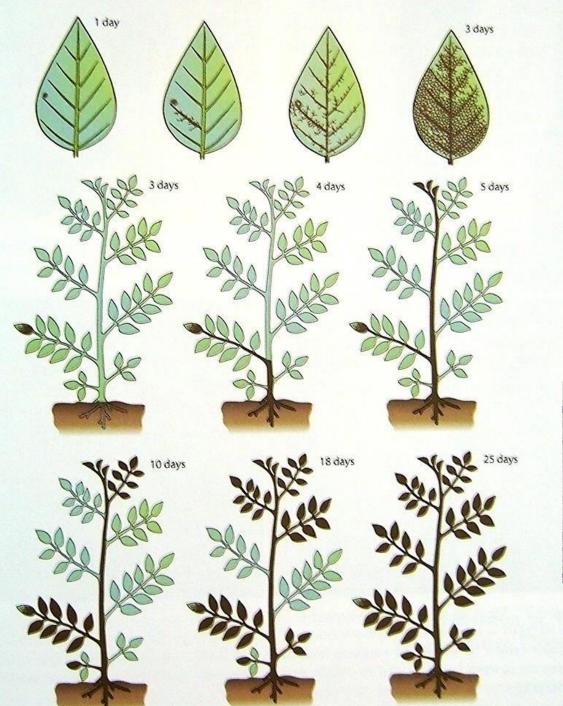




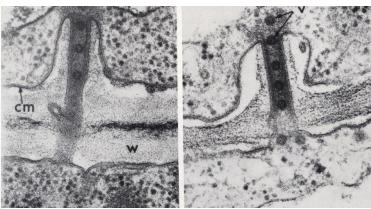
carborundum

Mechanical inoculation

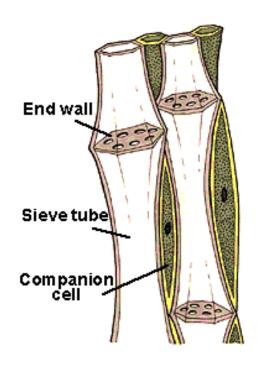




- Movement of virus in plant
- •8-10 mm/day by cell-to-cell movement

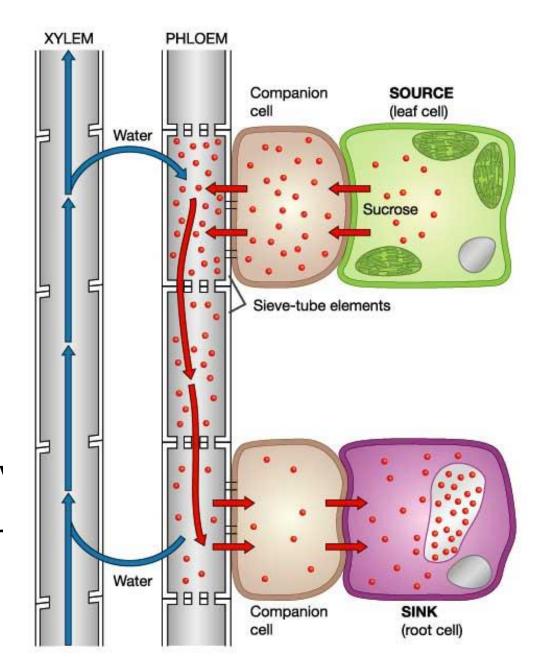


Zinnia leafdahlia mosaic virus



Phloem cells are alive.

Companion cells provide the energy for the tube cells. The end walls of the tube cells have pores through which food is transported from cell to cell ir the form of dissolved sugars







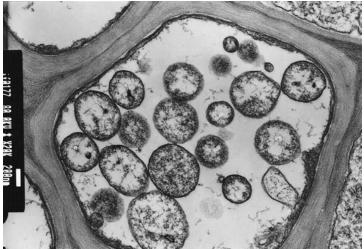




FIGURE 1 - Little leaf symptom on naturally infected bitter gourd plants exhibiting yellowish green, thickened, puckered leaves and internodes shortening (white arrow) compared to normal healthy vine (black arrow).

DISEASE DETECTION & IDENTIFICATION

Diagnosis of Plant Viruses

- Observation of symptom on plant; leaf, flower, fruit, stem, root, tuber, bulb, etc.
- Data collection on field area and environment
- Observation of signs that may be caused by other plant pathogens (fungi, bacteria, nematode, nutrient deficiency, etc.) or insects.
- Identification of viruses

Identification of viruses

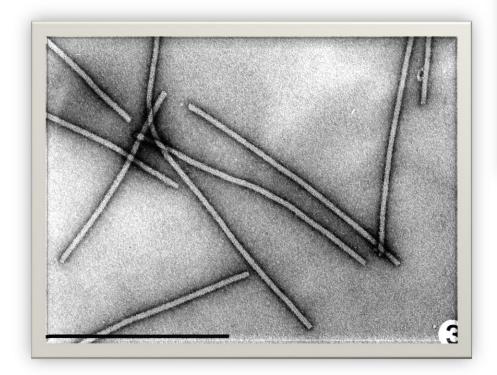
- Determination of size and shape by transmission electron microscope
- Determination of host range
- Determination of insect vectors
- Identification by serological techniques
 - Enzyme-linked immunosorbent assay (ELISA)
 - Dot immunobinding assay (DIBA)
 - Immunostrip, lateral flow assay
- Identification by DNA techniques
 - polymerase chain reaction (PCR)
 - DNA hybridization

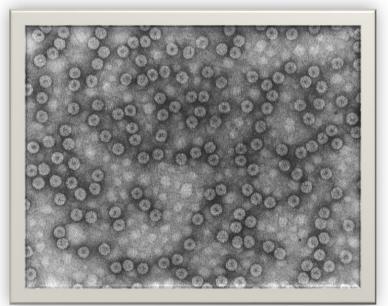
Determination of size and shape by transmission electron microscope



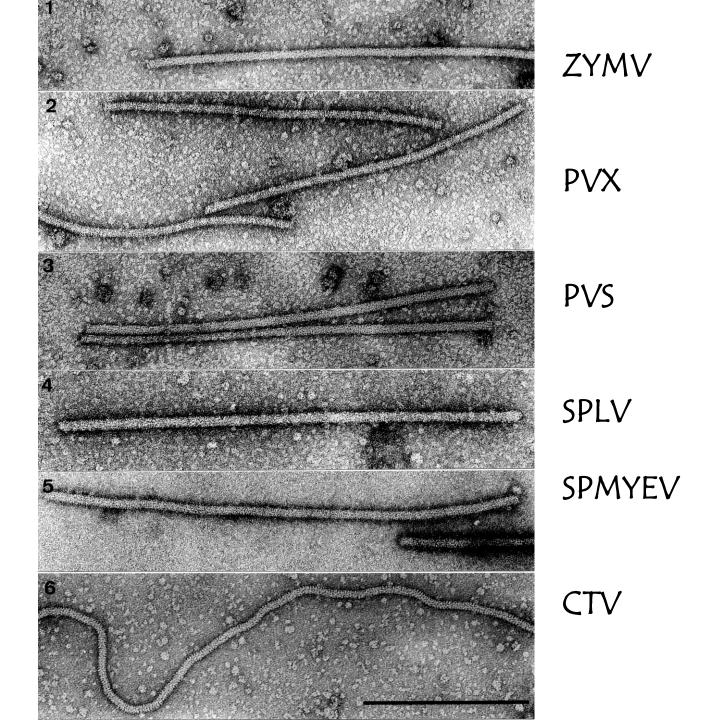


Morphology of plant viruses









Control of Virus Diseases

Prevention

- Virus-free propagating materials
- Resistant varieties; breeding, GMO
- Control of insect vectors
- Cross protection

Eradication

- Remove infected plants
- Eradicate weeds and alternative hosts
- Cultural practice; crop-free period, crop rotation...

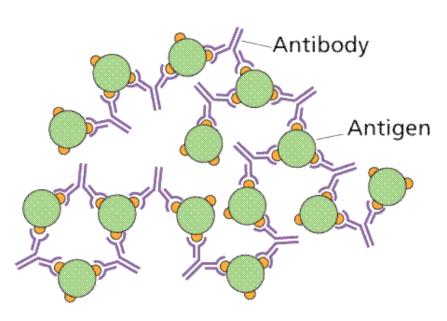
Therapy

Routine detection

- Identification by serological techniques
 - Enzyme-linked immunosorbent assay (ELISA)
 - Dot immunobinding assay (DIBA)
 - Immunostrip, lateral flow assay
- Identification by DNA techniques
 - Polymerase chain reaction (PCR)

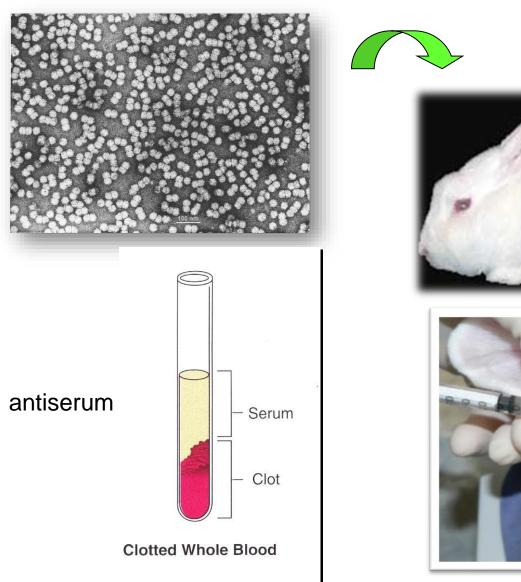
Identification by serological techniques





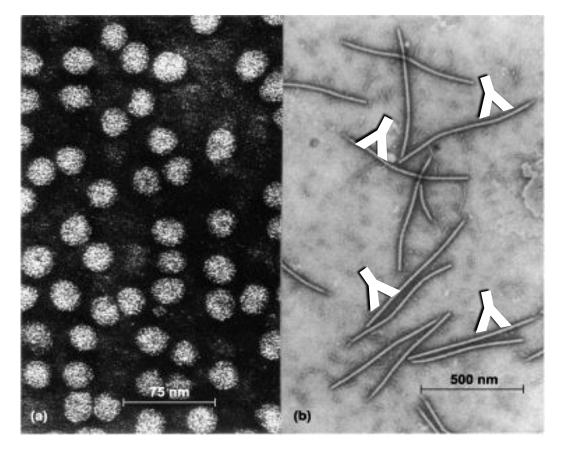
Antigen-antibody reaction

Antiserum production







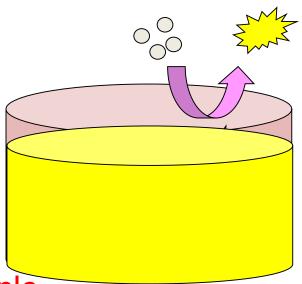


Virus-specific antibody

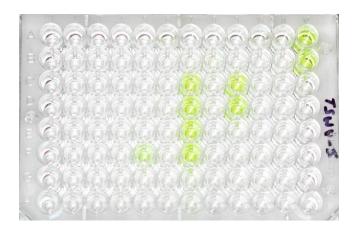
- Enzyme-linked immunosorbent assay (ELISA)
- Dot immunobinding assay (DIBA)
- Direct tissue blot immunoassay

Enzyme-linked immunosorbent assay (ELISA)





Plant sample





ELISA reader

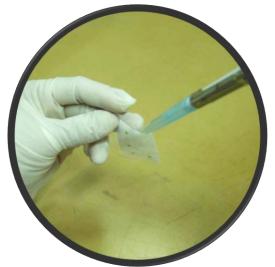


ELISA kit

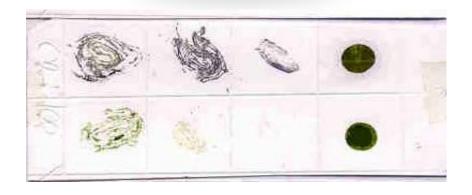
DOT IMMUNOBINDING ASSAY (DIBA)



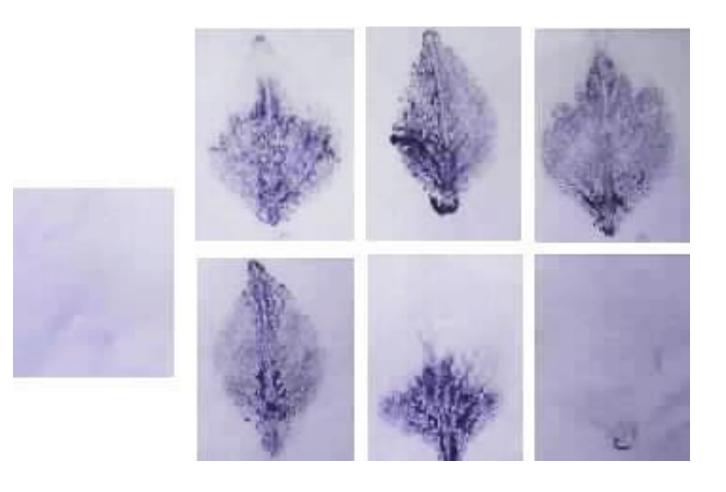
Nitrocellulose membrane Load drops of antigen (plant sap)



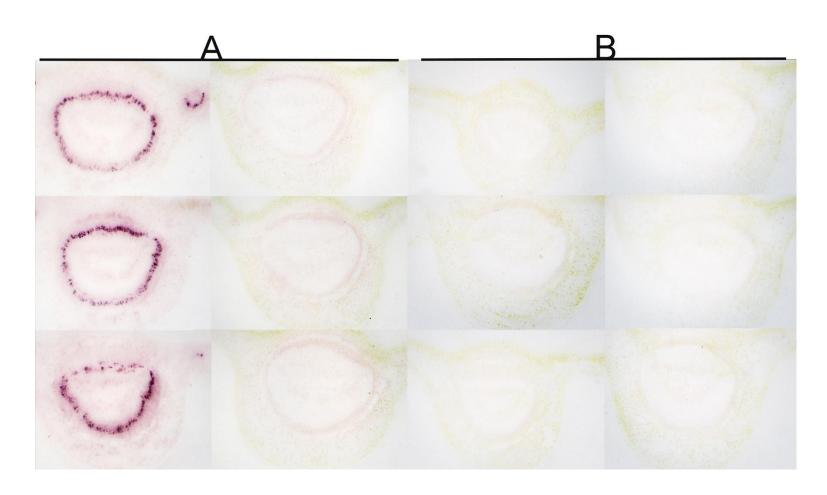
Reacting with specific antibody and color development in a plastic bag



Direct tissue blotting assay



Localization and Distribution of 'Candidatus Liberibacter asiaticus' in Citrus and Periwinkle by Direct Tissue Blot Immuno Assay with an Anti-OmpA Polyclonal Antibody

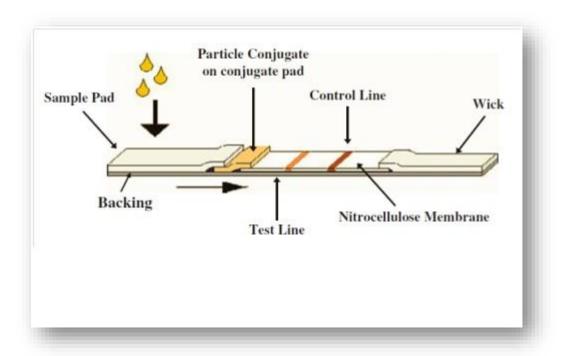




Positive Test Test line Negative Test (arrows point into test tube) Control line

Immunostrip (Lateral flow assay)

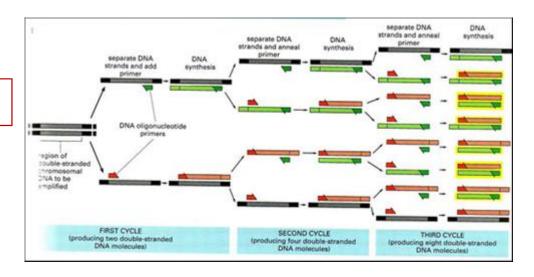






POLYMERASE CHAIN REACTION

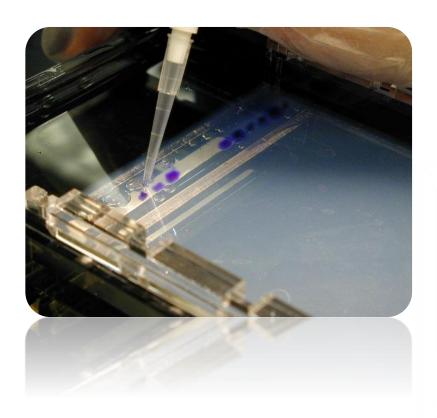
Specific primer for each virus



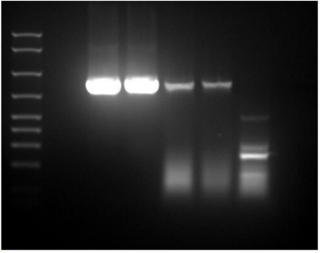












- M) Amplisize MW Marker
- 1-2) Control Plasmid
- 3-4) RT-PCR of CAII5) Beta Actin Control

Several PCR reactions were set up on the RNA isolated from experiment 14. One PCR reaction was a positive control using a plasmid with the CAII gene already cloned into it. The second reaction was an RT-PCR (reverse transcription - polymerase chain reaction) using the RNA isolated from experiment 13 to amplify the CAII gene. The final reaction was a beta actin control RT-PCR on the isolated RNA. Each reaction was set up in duplicate. An aliquot of each was loaded and run in a 1% agarose/TBE gel and stained with ethidium bromide.

Control of Virus Diseases

Prevention

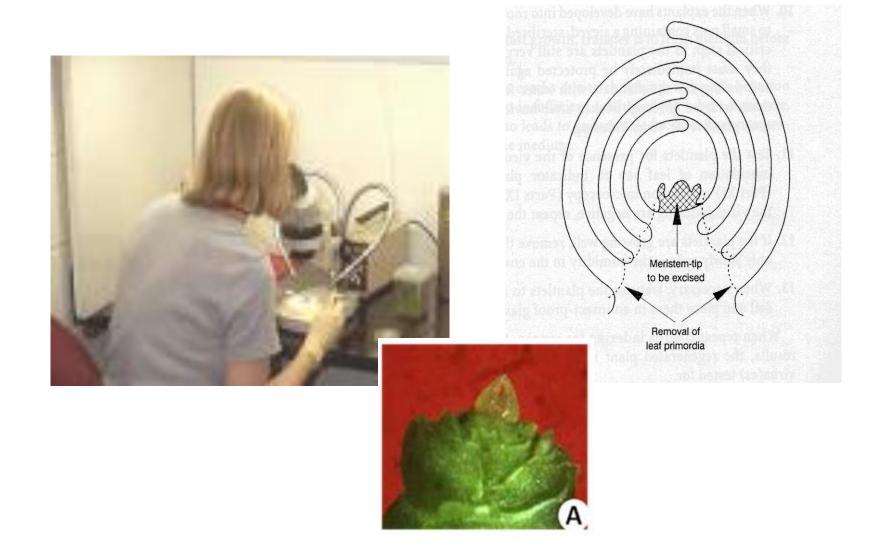
- Virus-free propagating materials
- Cross protection
- Resistant varieties; breeding, GMO
- Control of insect vectors

Eradication

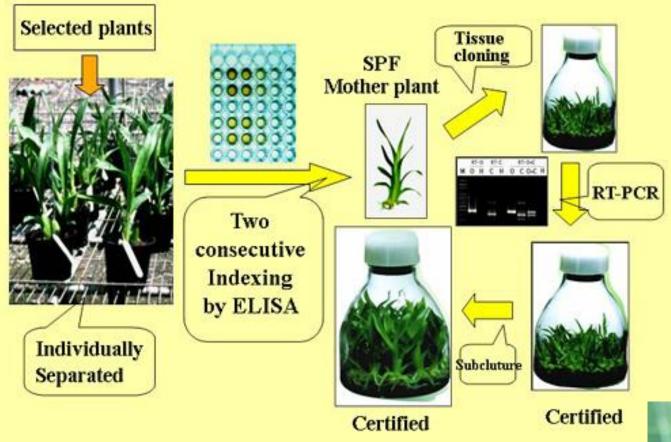
- Remove infected plants
- Eradicate weeds and alternative hosts
- Cultural practice; crop-free period, crop rotation...

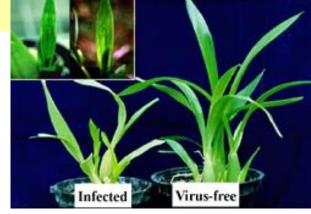
Therapy

Meristem-tip culture



Virus-free Oncidium seedling propagation and certification system

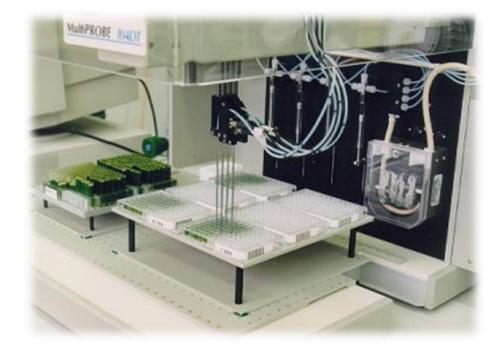




Certification of disease-free planting materials









Cross protection

by infecting young plants with mild strain of virus to prevent the infection of severe strain of the same virus.



Inoculate mild strain



No symptom after challenging with the severe strain



Resistant plant

Resistant varieties



Papaya resistant to papaya ring spot virus

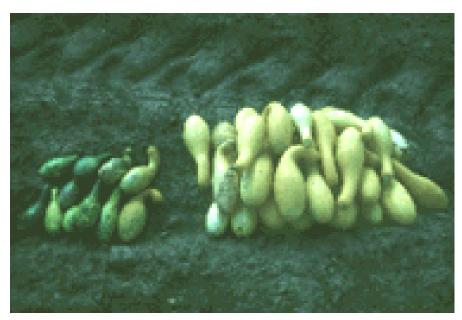


Squash resistant to...

Zucchini yellow mosaic virus (ZYMV),
 Watermelon mottle virus 2 (WMV-2), and
 Cucumber mosaic virus (CMV).







Comparative yield of fruit from border row (infected) with that from transgenic plant row that did not become infected.

squash zucchini yellow mosaic virus watermelon mosaic vurs –2



Effect of virus on fruit distortion and coloration. Fruits in the foreground are from virus infected plants, are distorted, and have mosaic green and yellow patterns. The fruits in the background are from the same field but are from virus resistant transgenic plants. Note that the fruits are not deformed and show the typical yellow color.

Control of Virus Diseases

- Prevention
 - Virus-free propagating materials
 - Resistant varieties; breeding, GMO
 - Control of insect vectors
 - Cross protection
- Eradication
 - Remove infected plants
 - Eradicate weeds and alternative hosts
 - Cultural practice; crop-free period, crop rotation...
- Therapy



Vegetable MD Online

Department of Plant Pathology, Ithaca, NY 14853

Diseases By Crop (Fact Sheets)

Search Function

Photo Gallery

News Articles/ Disease Alerts

Diagnostic Keys

Virus Weed Hosts/ Rotation Lists

Resistant Varieties

Glossary of Plant Pathology Terms

Vegetable Guidelines

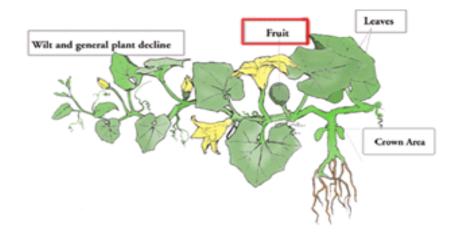
Vegetable IPM Links

Other Vegetable Links

Cornell Plant Disease Clinic

Home

Diagnostic Key:Diseases of Cucurbit Fruits



Bacteria

- Angular leaf spot
- Bacterial leaf spot

Phytoplasma

Aster Yellows

Viruses

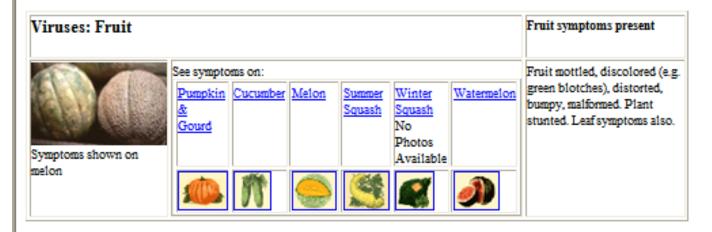
Viruses

Fungi/oomycete

- Anthracnose
- Choanephora
- Fusarium dry rot
- . Gummy stem blight black
- Phytophthora fruit rot
- Plectosporium blight
- Pythium
- Scab
- Sclerotinia white mold
- Septoria

Miscellaneous

Viruses



Miscellaneous



ALSO SEE:

Diagnostic Key -Cucurbit: Wilt and General Plant Decline and Crown Area

Diagnostic Key - Cucurbit: Diseases of Leaves



Vegetable MD Online

Diseases By Cron (Fact Sheet) Search Function Photo Gallery

Diagnostic Keys 2501 Virus Weed Hosts' Rotation Lists

Vegetable Guidelines

Veretable IFM Links Other Veretable Links

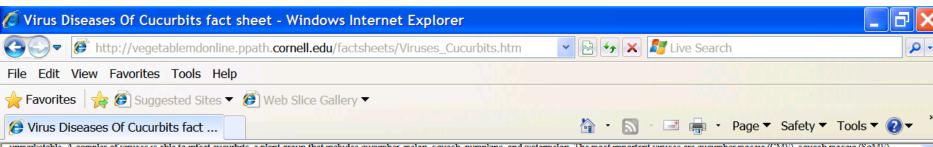
Cornell Flast Disease

Symptoms of viruses on watermelon fruit









unmarketable. A complex of viruses is able to infect cucurbits, a plant group that includes cucumber, melon, squash, pumpkins, and watermelon. The most important viruses are cucumber mosaic (CMV), squash mosaic (SqMV), watermelon mosaic I (WMV-1), watermelon mosaic 2 (WMV-2), and zucchini yellow mosaic (ZYMV). With the exception of SqMV, which is seedborne in melon and transmitted by beetles, the other major viruses are transmitted by several aphid species in a nonpersistent manner.

Major Cucurbit Viruses

Squash mosaic virus (SqMV) can cause an important disease of melons and squash in New York. The virus is seedborne in muskmelon and is spread in nature principally by the spotted and striped cucumber beetles. The virus is carried within the seed and cannot be eliminated by hot water or chemical treatment with tusodium phosphate.

Symptoms consist of pronounced chlorotic mottle, green veinbanding, and distortion of leaves of young seedlings. On mature plants, leaves show intense dark green mosaic, blistering, and hardening, suggestive of a hormonal herbicide effect (fig. 1). Infected fruit coming from such plants show a strong mottled pattern with a lack of netting on melons (fig. 2). Control measures include selection of disease-free seed and cucumber beetle control.

Cucumber mosaic virus (CMV) is probably the most widely distributed and important virus disease of cucurbits in New York. The virus overwinters in many perennial weed sources especially attractive to aphids when weed growth resumes in the spring. Early infection of squash and melons is particularly common. Aphids are the main and most efficient method of virus spread. Summer squash displays severe downward cupping along the midvein and leaf reduction from which the plants fail to recover (fig. 3). Color breaking of squash fruit is usually seen, but is not unique for this virus; other viruses causing this symptom include watermelon mosaic viruses 1 and 2, squash mosaic virus, and zucchini yellow mosaic virus. Early decline of muskmelon vines is usually attributed to CMV infection and should not be confused with collapse or "sudden wilt," which is a more complex disease and a plant-stress-related syndrome. CMV may be seedbome to a limited extent in some crops and weeds such as common chickweed (Stellaria media). Good CMV-resistant (actually tolerant since plants are infected by the virus) cucumber varieties are commercially available and produce a high percentage of unmottled fruit. All other commercially grown cucurbits are susceptible to CMV, although in yellow summer squash varieties that also carry a "precocious yellow gene," this gene serves to mask the color breaking common with cucurbit viruses (see discussion under WMV-2).

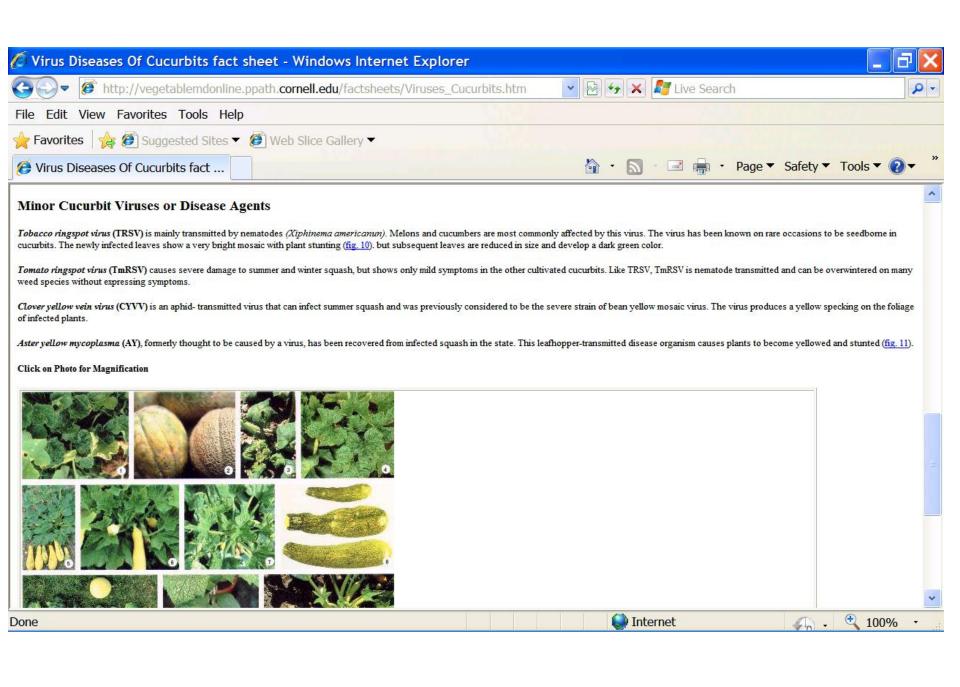
Watermelon mosaic virus 2 (WMV-2) is the second most important cucurbit virus in New York. This virus can infect and produce symptoms on all commercially grown cucurbits. This aphid-transmitted virus causes milder symptoms on the foliage of most infected plants like squash (fig. 4), and growers have seen a lessening of foliar symptoms following fertilization. Fruit distortion and color breaking are still a problem on varieties like yellow straightneck squash (fig. 5). Use of varieties such as 'Multipik' (fig. 6) can prolong the harvest period because the fruit are marketable in spite of foliar symptoms. The host range for WMV-2 is not limited to cucurbits, thus opening the possible overwintering of this virus in several leguminous species such as clover. Mixed infections of cucurbits with CMV and WMV-2 are common by the end of the season.

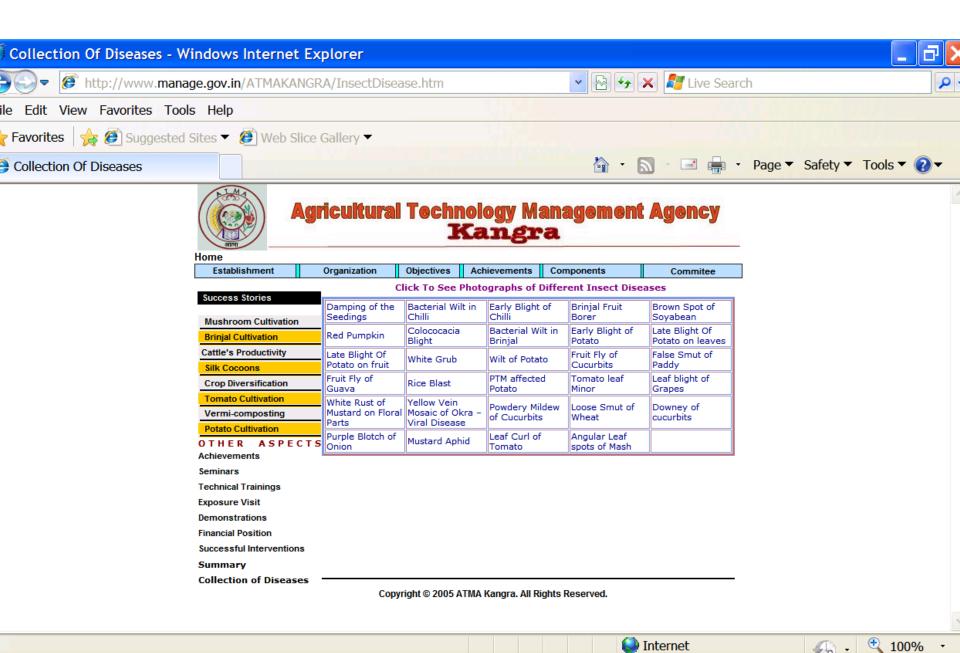
Watermelon mosaic virus 1 (WMV-1) is aphid transmitted, and infection is limited to cucurbits. Although more common in the southern and western regions of the United States, this virus has been recovered in New York several times since it first occurred in epidemic proportions in 1969. This virus is capable of infecting all commercial cucurbit crops. The foliage of affected plants shows strong mosaic, distortion, and deep leaf serration (fig. 7). Fruits are also malformed with knobby overgrowth (fig. 8).

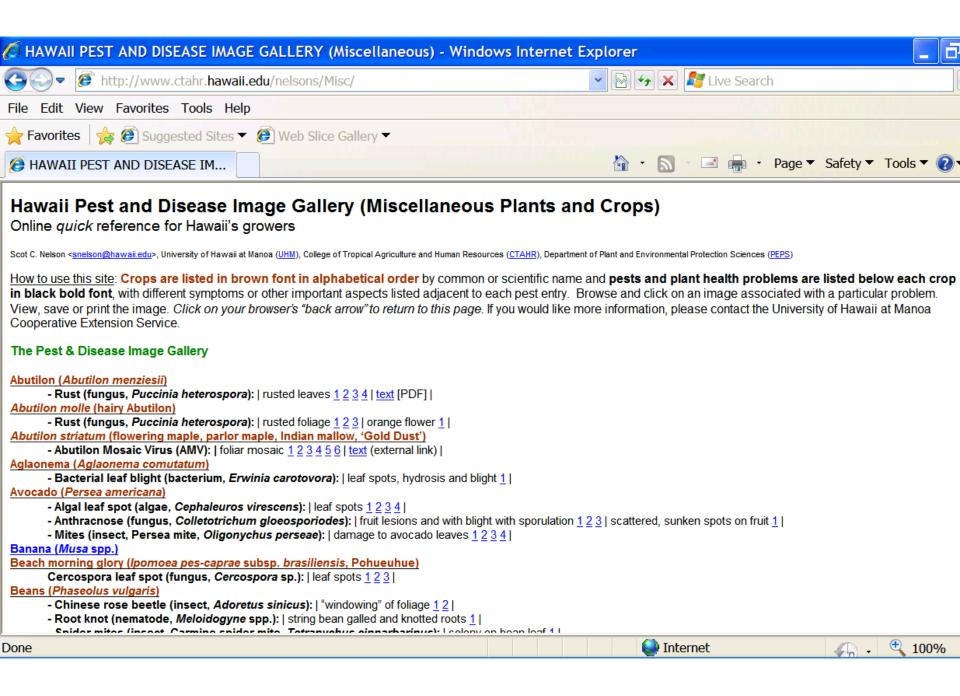
Zucchini yellow mosaic virus (ZYMV) is a recently described virus disease of cucurbits, first identified in Europe in 1981. It has since been reported from most southern and southwestern states and was found in New York State in 1983. The virus has characteristics very similar to WMV-1 and WMV-2 (nonpersistent aphid transmission, etc.), and like WMV-2, its host range is not limited to cucurbits. Currently, none of the genetic factors that confer resistance to WMV-1 or WMV-2 are able to control ZYMV, but other resistance sources have been identified. Muskmelon, watermelon, and squash are severely affected by ZYMV. Foliar symptoms consist of a prominent yellow mosaic, necrosis, distortion, and stunting. Fruits remain small, greatly malformed, and green mottled, including fruit of the variety 'Multipik' (fig. 9). It is too early to tell which weed hosts may serve to overwinter this virus in New York.

Minor Cucurbit Viruses or Disease Agents











Tom yum koong



Som tum



Massaman Beef Curry

